

ANTI-PROLIFERATIVE EFFECT OF METHYL GALLATE ISOLATED FROM
M. Pajang Kosterman IN SELECTED CANCER CELL LINES

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ABSTRACT

Mangifera pajang (Bambangan) is an indigenous fruit originated from Borneo Island. The kernel of *M. pajang* has been reported to possess various health benefits due to the phytochemicals content. As cancer is one of the most leading cause of death in Malaysia, this study was aimed to isolate and elucidate an aromatic ester, methyl gallate from the kernel extract of the *M. pajang* and determine the anti-proliferative potential of aromatic ester, methyl gallate which induce the cell cycle arrest and killing mechanism via apoptosis in selected cancer cell line. Chromatography methods were used in this study to isolate methyl gallate ($C_8H_8O_5$) from methanol crude extract. Identification of the isolated compound was done by spectroscopic methods including infrared (IR), mass spectrometry (MS), nuclear magnetic resonance (NMR) and comparison with reported data. The potential of anti-proliferative activity was investigated by using 3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay against breast (MCF-7 and MDA-MB-231), prostate (PC-3), pancreatic (Capan-2) and colon (HCT-116) cancer cell lines. The effect of methyl gallate on cell cycle arrest and apoptosis were confirmed by using cell cycle analysis and Annexin V staining. The results showed that methyl gallate displayed significant anti-proliferative activity against MCF-7 and moderate activity against PC-3 with an IC_{50} values of 54.7 μM and 97.6 μM respectively. Based on the IC_{50} value of MTT assay, MCF-7 cell line was selected for further determination of cell cycle arrest and apoptosis analysis. The cell cycle progression of MCF-7 cells treated with methyl gallate showed increased in cell population in sub- G_1 phase indicating apoptosis and cell cycle arrest at G_0/G_1 . In addition, the effect of methyl gallate on apoptosis induction resulted in total apoptotic cells were increased in time dependent manner. As a conclusion, methyl gallate isolated from kernel of *M. pajang* possess antiproliferative effect against breast cancer cell via apoptosis pathway.

ABSTRAK

Mangifera pajang (Bambangan) ialah buah-buahan nadir yang berasal dari kepulauan Boneo. Biji buah *M. pajang* dilaporkan mempunyai pelbagai manfaat terhadap kesihatan dan ini adalah disebabkan kehadiran pelbagai fitokimia. Memandangkan kanser merupakan penyebab utama kematian di Malaysia, objektif kajian ini adalah untuk memencilkan sebatian ester aromatik, metil gallat ($C_8H_8O_5$) daripada ekstrak biji buah *M. pajang* dan menentukan potensi aktiviti anti-proliferatif yang telah mempengaruhi mekanisme pembunuhan sel secara terkawal (apoptosis) dan fasa-fasa sel. Kaedah kromatografi yang digunakan telah berjaya memencilkan ester aromatik, metil gallat ($C_8H_8O_5$) daripada ekstrak mentah metanol. Pengenalpastian sebatian dilakukan dengan menggunakan kaedah spektroskopi termasuk inframerah (IR), spektroskopi jisim (MS), resonans magnetik nuklear (NMR) dan perbandingan data yang sedia ada. Potensi aktiviti anti-proliferatif oleh sebatian pencilan telah dilakukan dengan menggunakan asai MTT terhadap beberapa jenis sel kanser seperti kanser payudara (MCF-7 dan MDA-MB-231), prostat (PC-3), pankreas (Capan-2), dan kolon (HCT-116). Analisis pembunuhan sel yang dirawat dengan metil gallat terhadap kematian secara apoptosis dan fasa-fasa sel dilakukan dengan analisis fasa sel dan pewarnaan Annexin V. Keputusan aktiviti sitotoksik oleh metil gallat menunjukkan aktiviti yang signifikan terhadap sel kanser payudara (MCF-7) dengan dengan nilai IC_{50} 54.7 μ M dan aktiviti sederhana terhadap sel kanser prostat (PC-3) dengan nilai IC_{50} 97.6 μ M. Berdasarkan keputusan asai MTT, sel kanser payudara (MCF-7) telah dipilih untuk kajian mekanisme pembunuhan sel dan fasa sel. MCF-7 sel yang dirawat dengan metil gallat menunjukkan peningkatan peratusan sel dalam fasa sub- G_0 dan terdapat penahanan sel berlaku pada fasa G_0/G_1 . Manakala, keputusan asai apoptosis menunjukkan sel yang dirawat dengan metil gallat mendorong kematian secara selari dengan masa rawatan. Sebagai rumusan, sebatian terpencil daripada biji buah *M. pajang*, metil gallat mempunyai kesan sebagai bahan anti-proliferatif ke atas sel barah payudara.

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LIST OF SYMBOLS AND ABBREVIATIONS

°C	-	Celcius
cm	-	Centimeter
g	-	Gram
h	-	Hour
IC ₅₀	-	Concentration that needed to produce 50% cells inhibition
L	-	Litre
M	-	Molar
mg	-	miligram
mL	-	mililitre
mm	-	milimeter
mM	-	miliMolar
nm	-	nanometer
µg	-	microgram
µL	-	microlitre
µM	-	micro molar
ATCC	-	American Type Culture Collection
CC	-	Column chromatography
CDK	-	Cyclin Dependent Kinase
DIP	-	Direct Induction Probe
DNA	-	Deoxyribonucleic acid
DPPH	-	2,2-diphenyl-1-ptycrylhydrazyl radical
EI-MS	-	Electron Impact-Mass Spectrometry
FBS	-	Fetal Bovine Serum
FRAP	-	Ferric reducing/ antioxidant power
FT-IR	-	Fourier Transform-Infrared
G ₀	-	Resting phase

G ₁	-	Gap 1 phase
G ₂	-	Gap 2 phase
HCT-116	-	Colon Cancer cell lines
MCF-7	-	Hormone dependent breast carcinoma cell line
MCF-10A	-	Normal human breast cell
MDA-MB-231	-	Non-hormone dependent breast carcinoma cell line
MHz	-	MegaHertz
MOH	-	Ministry of Health
MTT	-	3-(4,5-dimethylthiazol-2-yl)-2,5-diphenyltetrazolium bromide
NMR	-	Nuclear Magnetic Resonance
PC-3	-	Prostate carcinoma cell line
PI	-	Propidium Iodide
RNase	-	Ribonuclease
RPMI 1640	-	Roswell Park Memorial Institute 1640
S	-	Synthesis phase
TFC	-	Total Flavonoid Content
TLC	-	Thin Layer Chromatography
TPC	-	Total Phenolic Content
WHO	-	World Health Organization

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Journals:

- (i) **Azlen Che Rahim**, Mohd Fadzelly Abu Bakar, Nur Kartinee Kassim, Johnson Stanslas, Wan Nor Izzah Wan Mohamad Zain. (2018). Selective cytotoxic activity of methyl 3,4,5-trihydroxybenzoate isolated from Kernel of Bambangan (*Mangifera pajang*). *Asian Journal of Chemistry*, 30(10): 2273-2276.
- (ii) **Azlen Che Rahim**, Mohd Fadzelly Abu Bakar, Nur Kartinee Kassim, Johnson Stanslas. Methyl Gallate isolated from *Mangifera pajang* display anti-cancer activity by induction of cell arrest and apoptosis in human breast cancer cell. [Submitted to *Food and Function*]

Conference:

- (i) International Conference on Drug Discovery and Translational Medicine (ICDDTM 2018), 4-5 December 2018, The Everly Hotel, Putrajaya Malaysia.

CHAPTER 1

INTRODUCTION

1.1 Background of Research

Cancer is a major public health burden worldwide and it is the leading cause of death in both developed and developing countries. The increasing number of death due to cancer is correlated with the modernisation of lifestyle, improved socioeconomic status (Asegaonkar *et al.*, 2015), and exposure to sunlight (Zima *et al.*, 2001). According to a report by Torre *et al.* (2015), high cancer incidences including lung, colorectal, breast, and prostate cancer are reported to occur at high income countries (HIC). Meanwhile, cancer incidence for low and middle-income countries (LMIC) is growing, with stomach, liver, oesophageal, and cervical cancer having the highest rate. It is also estimated that over 20 million of new cases will be expected annually by as early as 2025 (Bray, 2014). According to an online database of The International Agency for Research on Cancer (IARC)- GLOBOCAN 2012, prostate and lung cancer are top cases commonly diagnosed among males in 87 countries worldwide, while breast and cervical cancers are the most frequently diagnosed in females (Torre *et al.*, 2015).

Chemoprevention has become familiar in the cancer research area, particularly in drug development, pharmacology, and molecular biology research. People has realised that effective cancer treatments such as chemotherapy, radiotherapy, and drug therapy still could not challenge the potential of chemoprevention as the survival rate of cancer patients is still low (Ladas, 2004; Liu, 2009). Chemoprevention is an alternative in preventing the development of cancer by using specific agents either to inhibit, delay, or reverse carcinogenesis (Tamimi *et al.*, 2002). Meanwhile, chemotherapy is another term that is involved in the cancer research. Unlike the

chemoprevention, chemotherapy is one of modes of treatment in cancer patients with the utilisation of chemotherapeutic drugs (Liu, 2009).

Malaysia is one of the Asian countries that are blessed with diverse biological resources and high in percentage of flora, which are believed to have medicinal and nutritional values. It is estimated that there are about 370 species of local fruits in Malaysia (Rukayah, 2002; Mirfat, Salma, & Razali, 2016). Other than for commercialisation purposes, tropical fruits in Malaysia are grown for local consumption and medicinal uses. Most of the indigenous fruits in Malaysia are wild fruits that are naturally distributed in the forests and most of them are underutilised (Abu Bakar & Fry, 2013). Some of the underutilised fruits especially in Borneo that are available in Malaysia are bacang (*Mangifera foetida*), bambangan (*Mangifera pajang*), cerapu (*Garcinia prainiana*), durian (*Durio*), jambu (*Syzygium aqueum*), kuini (*Mangifera odorata*), pulasan (*Nephelium mutabile*), and salak (*Salacca zalacca*) (Hock *et al.*, 2016). Many of these fruits are unpopular due to the lack of promotion, not being fully explored, and minimal space for planting (Chai *et al.*, 2008).

The nutritional and medicinal benefits of plants, fruits, and vegetables are receiving great attention nowadays as they have been shown to provide a better support for human wellbeing. Recently, plant-derived products have been the major target for many studies due to their versatile applications. Some medicinal plants have been reported to possess anticancer activity, where polyphenols from fruits and vegetables are the responsible molecules for chemopreventive effects (Mutalib *et al.*, 2016). It has also been reported that plant-derived materials possess highly diverse and complex molecular structures compared to synthetic drugs and play an important role in human health and in the development of new anticancer drugs (Faezizadeh, Gharib, & Godarzee, 2016). A study by Liu (2004) also reported that diets which are rich in fruits and vegetables are good sources of natural phytochemicals which work synergistically to exhibit antioxidant and anticancer activities.

Phytochemicals in plants play a major role in contributing to human health. High dietary intake of fruits and vegetables as well as whole grains is strongly associated with reduced risk of getting chronic diseases, such as cancer and cardiovascular diseases (CVD). The United States National Academy of Sciences also emphasised the importance of adding citrus fruits, carotene-rich fruits, and vegetables to diet to reduce the risk of cancer (Liu, 2003). Research and development of rare and underutilised fruits is becoming economically important nowadays as it promotes a

new generation of ‘superfruits’ which have the potential to be developed as functional food and nutraceuticals.

In Southeast Asia, there are many varieties of local fruits such as mango (*Mangifera indica*), durian (*Durio*), rambutan (*Nephelium lappaceum*), and papaya (*Carica papaya*), which are sold locally and globally. However, there are also fruits that are sold locally but have not been fully explored. For example, *Mangifera pajang* is a native fruit that can only be found in the Borneo Islands (Malaysia: Sabah, Sarawak; Brunei; Indonesia: Kalimantan) and is rarely discovered in Peninsular Malaysia. This fruit is prevalent among the Kadazan-Dusun community in Sabah as an accompaniment to their meal and traditional cuisine (Tangah *et al.*, 2017). Many studies on *M. pajang* phytochemicals have reported their health-beneficial properties.

1.2 Problem statement

Based on cancer statistics, deaths caused by cancers are known to increase annually. The leading cancers among males in Malaysia are lung, nasopharynx, stomach, urinary bladder, rectum, liver, and colon cancers. Meanwhile, cervix, breast, lung, colon, and rectum cancers are majorly being diagnosed in females in Malaysia (Kasri, 1993). In addition, Azizah *et al.* (2016) added that Malay females have the highest rate of breast cancer incidence. The increase in death rate due to cancer are related to several factors such as burden to patients due to the increase in cost for cancer treatment. Not only that, current cancer therapies including surgery, radiotherapy, chemotherapy, and drugs have a major drawback towards the patients such as bad side effects after the therapy (Partridge *et al.*, 2001). The cancer patients must bear with symptoms such as nausea, anaemia, and hair loss. Not only that, the development of chemoresistance is a persistent problem and makes it difficult for the patients to deal with the treatment. There is thus a great need towards finding a better alternative on cancer therapy including traditional medicine, which has become an important approach to control cancer. Apart from that, there has been a focus on natural products from plant sources to be developed as functional food to provide better ways for cancer patients to reduce their burden.

1.3 Objectives of the study

General objective:

To explore the potential of local fruits, *Mangifera pajang* to be developed as nutritional food.

Specific objectives:

- (i) To isolate and elucidate methyl gallate from the kernel of *M. pajang*.
- (ii) To evaluate the *in-vitro* anti-proliferative activity of the methyl gallate extracted from kernel extract of *M. pajang* against selected cancer cell lines
- (iii) To evaluate the effect of methyl gallate isolated from kernel extract of *M. pajang* on apoptosis event and cell cycle arrest in selected cancer cell lines.

1.4 Significance of study

With the increasing trend of cancer incidence of each year, number of efforts are needed in order to provide better alternative to reduce the percentage of mortality due to cancer. One of the efforts is through the changes in diet (Willet, 1995). Diet that is rich in plants and fruits could be a better solution in reducing cancer incidence as fruits and plants are known to possess various health benefits and anticancer properties. Fruits and vegetables play a vital role in the health of human beings by providing various phytochemicals such as phenolic compounds and flavonoids to promote health (Abu Bakar *et al.*, 2010). These compounds are known to be natural antioxidants which are abundantly present in plants and fruits. These compounds have been the main target for scientists as they provide not only nutritional value but have promising effects on health as well.

Since many studies have provided findings on the biological and health-promoting effects of *M. pajang*, further evaluation of anticancer properties of the active extract from this fruit can provide more details about the active compounds and their biological activities. With the aim to identify and isolate active natural compounds from the active extract of *M. pajang* fruit, this study provides a better explanation in the fundamental mechanism of actions in killing cancer cells.

1.5 Scope of study

In the present study, the antiproliferative potentials of natural products were explored. *M. pajang* fruit is abundantly found in Borneo Island and the focus was given on its kernel part. Given that many past studies have reported that *M. pajang* kernel displayed many health benefits such as antioxidant activity due to the diversity of polyphenols. Higher antioxidant activity has suggested by some in-vitro and in-vivo study which certain antioxidant agent selectively inhibit the growth of tumour cells, induce the cellular differentiation, and also may alter the intracellular redox state, hence enhances the effect of cytotoxic therapy (Ladas *et al.*, 2004). In this study, emphases are given to the one of the remarks on the cancer manifestation that involved in the cancer development, which is apoptosis. The effects of the natural products on cancer development are also discussed.



CHAPTER 2

LITERATURE REVIEW

2.1 Cancer

Cancer is a disease where abnormal cells continue to grow in an uncontrolled manner and disregard the normal rules of cell division (Kumar *et al.*, 2016). Normally, body cells control the process of cell division, cell death, and cell differentiation, but cancer cells ignore the regulation, resulting in uncontrolled cell growth and proliferation (Li & Wang, 2014). These uncontrolled cancer cells will proliferate, resulting in the formation of tumours (Kumar *et al.*, 2016). This abnormal proliferation can continue and spread to other parts of the body; this is called metastasis and may be fatal (Hejmadi, 2009).

There are two types of cancer cells, which are malignant and benign tumour cells. Benign tumours are tumours that do not spread and usually pose little threat to the host since they are localised and small (Lodish, Berk, & Zipursky, 2000). On the other hand, malignant tumours are non-localised and invade other surrounding tissues (Lodish *et al.*, 2000). A benign tumour has a slower growth speed than a malignant one (Baba & Catoi, 2007).

Cancer can be treated in several ways; the available technologies today are surgery, radiotherapy, immunotherapy, and chemotherapy. Some of the methods use anticancer drugs or defined as chemopreventive agents, such as those derived from plants such as taxol (found in *Taxus brevifolia*), combretastatin A-4 (from *Combretum caffrum*) (Srivastava *et al.*, 2005), camptothecin (isolated from *Camptotheca acuminata*), vinblastine, and vincristine (found in *Catharanthus roseus*) (Colgate & Molyneux, 2007).

2.1.1 Apoptosis and cell cycle

Apoptosis is a programmed, physiological mode of cell death mechanism for removing unwanted and detrimental cells in a silent manner during embryonic development, tissue homeostasis, and immune regulation (Orangi *et al.*, 2016). The normal regulation of this molecular mechanism becomes a drawback when the pathway of apoptotic signalling is altered, leading to development of cancer and diseases (Favaloro *et al.*, 2012). Cells that have severe DNA damage and are unsuccessfully removed by apoptosis would acquire mutation and this enables them to grow and proliferate uncontrollably.

Apoptosis can be initiated by two pathways: extrinsic and intrinsic. The extrinsic pathway is induced by extracellular stimuli such as binding of ligand to death receptor on cell surface or cytokine stimulation which results in the activation of caspase cascade and alteration in gene expression profile. Meanwhile, the intrinsic pathway is induced by many ranges of stimuli such as DNA damage, transcription/translation damage, or virus infection (Hollier *et al.*, 2007).

Caspase-dependent apoptosis is characterised by the activation of a series of pathways, leading to the activation of a family of proteases (caspases), resulting in an ordered disruption, which is a more favourable way of cell disruption without the leakage of cellular components and induction of inflammation (Favaloro *et al.*, 2012).

Apoptosis is more favourable than necrosis due to the response of the cells towards receiving the cell-death signals. A report by Saraste & Pulkki (2000) briefly described the characteristics of cell at the onset of apoptosis: it begins with the shrinkage of the cell and the nucleus, followed by condensation of nuclear chromatin masses. Next, the nucleus progressively breaks up; this is termed as karyorrhexis. The budding cells are detached from the surrounding tissue and become extensions and the plasma membrane seals to form a separate membrane around detached solid cellular materials, or known as apoptotic bodies, which are enclosed together with cellular organelles and fragments of the nucleus. Later, these apoptotic bodies are engulfed by neighbouring cells such as macrophages and parenchyma cells (Kerr *et al.*, 1994).

In a cell cycle research, it is necessary to synchronise a population of cells at a stage in the cell cycle, so that the cellular or biochemical features of that stage can be analysed. Numerous drugs, including thymidine and hydroxyurea, block DNA synthesis by inhibiting the synthesis of specific nucleotides, which results in a

REFERENCES

- Abdul Aziz, N., Mohd Amin, F., Nik Farid, N. D., Dahlui, M. (2013). Breast Cancer awareness of Rural Women in Malaysia: is it the same as in the cities? *Asian Pacific Journal Cancer Prevention*. 14 (12): 7161-7164
- Abdul Rahim, M. S. A., Salihon, J., Mohd Yusoff, M., Martua Damanik, M. R. (2013). Antioxidative activity and phenols content in Five Tropical Lamiaceae Plants. *Journal of Tropical Resources and Sustainable Science*. 1(2): 49-54
- Abu Bakar, M. F., Mohamed, M. Rahmat, A., and Fry, J. R. (2009). Phytochemicals and Antioxidant activity of different parts of Bambang (Mangifera pajang) and tarap (Artocarpus odoratissimus), *Food Chemistry*, 113:479-483.
- Abu Bakar, M. F., Abdul Karim, F., Suleiman, M., Isha, A., Rahmat, A. (2015). Phytochemicals Constituents, Antioxidant and Antiproliferative properties of a Liverwort, *Lepidozia borneensis* Stephani from Mount Kinabalu, Sabah, Malaysia. *Evidence-Based Complementary and Alternative Medicine*.
- Abu Bakar, M. F., Mohamed, M., Rahmat, A., Burr, S. A., Fry, J. R., (2013). Cellular assessment of the extract of bambangan (*Mangifera pajang*) as a potential cytoprotective agent for the human hepatocellular HepG2 cell line. *Food Chemistry*.136:18-25
- Abu Bakar, M. F., Mohamed, M., Rahmat, A., Burr, S., A., and Fry, J. R. (2010). Cytotoxicity and polyphenol diversity in selected parts of *Mangifera pajang* and *Artocarpus odoratissimus* fruits. *Nutrition & Foods Science*. 40(1): 29-38.
- Abu Bakar, M. F., and Fry, J. R. (2013). A review on underutilized indigenous bambangan (*Mangifera pajang*) fruit as a potential novel source for functional food and medicine. 2013. *Journal of Medicinal Plants Research*. 7(45):3292-3297.
- Adan, A., Alizada, G., Kiraz, Y. Baran, Y. Nalbant, A., (2016). Flow Cytometry: Basic Principles and Applications. *Critical Reviews in Biotechnology*. 1-14. Doi:10.3109/07388551.2015.1128876

- Adom, K. K., Sorrells, M. E., and Liu, R. H. (2005). Phytochemicals and antioxidant activity of milled fractions of different wheat varieties. *Journal of Agricultural and Food Chemistry*. **53**(6): 2297-2306
- Ahmad, S., Sukari, M. A., Ismail, N., Ismail, I., S., Abdul, A. B., Abu Bakar, M. F., Kifli, N, (2015) Phytochemicals from *Mangifera pajang* Kosterman and their biological activities. *BioMed Central (BMC) Complementary & Alternative Medicine*. **15**:83. Doi: 10.1186/s12906-015-0594-7
- Ali I., Waseem, A., Saleem, K. (2011). Cancer scenario in India with future perspective. *Cancer Theraphy*, **8**, 56-32.
- Allred, D. C. (2010). Ductal carcinoma in situ; terminology, classification and natural history. *Journal of the National Cancer Institute*, **2010**(41): 134-138. Doi:10.1093/jncimonographs/lgq035
- Al-Sheraji, S. H., Ismail, A., Manap, M. Y., Mustafa, S., Yusof, R. M., & Hassan, F. A. (2012). Purification, characterization, and antioxidant activity of polysaccharides extracted from fibrous pulp of *Mangifera pajang* fruits. *Food Science and Technology*. **48**:291-296
- Althuis, M. D., Dozier J. M., Anderson, W. F., Devasa, S. S., Brinton, L. A., (2005). Global trends in breast cancer incidence and mortality 1973-1997. *International Journal of Epidemiology*, **34**: 405-12.
- Aman, R. (1999). Buah Nadir Malaysia. Dewan Bahasa dan Pustaka, Kuala Lumpur, Malaysia.
- Anderson, K. E., Mongin, S. J., Sinha, R., Stolzenberg-Solomon, R., Gross, M. D., Ziegler, R. G., Mabie, J. E., Risch, A., Kazin, S. S., Church, T. R. (2012). Pancreatic cancer risk: Associations with meat-derived carcinogen intake in the Prostate, Lung, Colorectal and Ovarian Cancer Screening Trial (PLCO) cohort. *Molecular Carcinogenesis*, **51**:128-137.
- Anderson, B. O., Cazap, E., El Saghir, N. S., Yip, C. H., Khaled, H. M., Otero, I. V., (2010). Optimisation of breast cancer management in low-resource and middle-resource countries: executive summary of Breast Health Global Initiative consensus. *Lancet Oncology* **2011**. **12**:387-98
- Appleby, P. N., Crowe, F. L., Bradbury, K. E., Travis, R. C., Key, T. J. (2016). Mortality in vegetarians and comparable nonvegetarians in the United Kingdom. *American Journal of Clinical Nutrition*, **103**:218-230

- Arnold, M., Sierra, M. S., Laversanne, M., Isabelle, S., Jemal, A., Bray, F. (2016). Global pattern and trends in colorectal cancer incidence and mortality. *Gut*, 66(4): 1-9
- Asegaonkar, S. B., Asegaonkar, B. N., Takalkar, U. V., Advani, S., Thorat, A. P. (2015). C-Reactive Protein and Breast Cancer: New Insights from old molecule. *International Journal of Breast Cancer*:1-6.
- Ascacio, J., Buenrostro, J., Aguilera, A., Prado, A., Rodriguez, R., Aguilar, C. (2011). Ellagitannins: Biosynthesis, biodegradation and biological properties. *Journal of Medicinal Plant Research*, 5(19), 4696-4703.
- Azizah, A., M., Norsaleha, I., Hashimah, A. N, Asmah, Z., & Mastulu, W., (2016). Malaysian National Cancer Registry Report 2007-2011. *National Cancer Institute*.
- Baba, A. I., & Catoi, C. (2007). Chapter 3, Tumor Cell Morphology. *Comparative Oncology*. Bucharest: The Publishing House of Romanian Academy.
- Bally, I. S. E., 2006. *Mangifera indica* (mango), Ver 3.1. In: Elevitch, C.R. (ed). Species Profiles for Pacific Island Agroforestry. Permanent Agriculture Resources (PAR), Hawaii. <http://www.traditionaltree.org>
- Bartel, B. (1997). Auxin biosynthesis. *Annual Review of Plant Physiology and Plant Molecule Biology*, 48: 51-66.
- Basset, G., Quinlivan, E., Ravanel, S., Rebeille, F., Nichols, B., Shinozaki, K., Seki, M., Adams-Phillips, L., Giovannoni, J., Gregory, J. and Hanson, A. (2004). Folate synthesis in plants: the p-aminobenzoate branch is initiated by a bifunctional PabB protein that is targeted to plastids. *Proceeding of the National Academic of United States*, 101:1496-1501.
- Bosetti, C., Bertuccio, P., Malvezzi, M., Levi, F., Chatenoud, L., Negri, E. Cancer mortality in Europe, 2005-2009 and an overview of trends since 1980. *Annals of Oncology*, 24:2657-71.
- Bostrom P. J. and Soloway, M. S. (2007). Secondary cancer after radiotherapy for prostate cancer: Should we be more aware of the risk?. *European Urology*, 52:973-982.
- Bray, F. (2014). Transitions in human development and the global cancer burden. *World Cancer Report*, 54-68.
- Brito, A. F., Ribeiro, M., Abrantes, A. M., Pires, A. S., Teixeira, R. J., Tralhao, J. G. and Botelho, M. F., (2015). Quercetin in Cancer Treatment, alone or in Combination

- with Conventional Therapeutics? *Current Medical Chemistry*, 22(26), 3025-3039.
- Center, M. M., Jemal, A., Lortet-Tieulent, J., Ward, E., Ferlay, J., Brawley, O. (2012). International variation in prostate cancer incidence and mortality rates. *European Urology*, 61:1079-92
- Chai, C. C., Teo, G. K., Lau, C. Y., Powoven, A. M. A., (2008). Conservation and sustainable utilization of indigenous vegetables of Sarawak. In: *Agrobiodiversity in Malaysia*, Kuala Lumpur, Malaysia. Pp.42-55.
- Chauduri, D., Ghate, N. B., Singh, S. S., Mandal, N. (2015). Methyl gallate isolated from *Spondias pinnata* exhibits anticancer activity against human glioblastoma by induction of apoptosis and sustained extracellular signal-regulated kinase $\frac{1}{2}$ activation. *Pharmacognosy Magazine*. 11(42):269-76.
- Chung, K. T., Wong, T. Y., Wei, C. I., Huang, Y. W., Lin, Y. (1998). Tannins and human health: A review. *Critical Reviews in Food Science and Nutrition*, 38(6): 421-64
- Chen, H. M., Wu, Y. C., Chia, Y. C., Chang, F. R., Hsu, H. K., Hsieh, Y. C. Chen, C. C., Yuan, S. S. (2009). Gallic acid, a major component of *Toona sinensis* leaf extracts, contains a ROS-mediated anti-cancer activity in human prostate cancer cells. *Cancer Letter*, 286:161-171.
- Chew, Y. L., Lim, Y. Y., Stanslas, J., Ee, G. C. L., Goh, J. K. 2014. Bioactivity guided isolation of anticancer agents from *Bauhinia kockian* Korth. *African Journal of Traditional, Complementary and Alternative Medicines*. 11(3):291-299.
- Chlebowski, R. T., Manson, J. E., Anderson, G. L., Cauley, J. A., Aragaki, A. K., Stefanick, M. L., (2013). Estrogen plus progestin and breast cancer incidence and mortality in the Women's health Initiative Observational Study. *Journal of the National Cancer Institute*. 105:526-35
- Cho, J. J., Chae, J., Yoon, G., Kim, K. H., Cho, J. H., Cho, S. S., Cho, Y. S., Shim, J. H., 2014. Licochalcone A, a natural chalconoid isolated from *Glycyrrhiza inflata* root, induces apoptosis via Sp1 and Sp1 regulatory proteins in oral squamous cell carcinoma. *International journal of oncology*. 45:667-674.
- Colditz, G. A., Baer, H. J., Tamimi, R. M., (2006). Breast Cancer. In: Schottenfeld D, Fraumeni J. F. Jr, editors. *Cancer epidemiology and prevention*. 3rd ed. New York: Oxford University Press, pp. 995-1012.

- Colgate, S. M. and Molyneux, R. J., (2007). Bioactive Natural Products: Detection, Isolation and Structural Determination. CRS Press: Florida.
- Croft, K. D. (1998). The chemistry and biological effects of flavonoids and phenolic acids. *Annals of the New York Academy of Sciences*. 854:435-42
- Crozier, A., Jaganath, I. B., Clifford, M. N. (2016). Phenol, polyphenols, and tannins: An Overview. *Plants Secondary Metabolites: Occurrence, Structure and role in human Diet*. 1-24. Doi: 10.1002/9780470988558.ch1
- Crispo, J. A., Piche, M., Ansell, D. R., Eibl, J. K., Tai, I. T., Kumar, A., Ross, G. M. and Tai, T. C. (2010). Protective effects of methyl gallate on H₂O₂- induced apoptosis in PC-12 cells. *Biochemical and Biophysical Research Communications*. 393:773-778.
- Da Silva, M. M., Iriguchi, E. K. K., Kassuya, C. A. L., Vieira, M. C., Fonglio, M. A., De Carvalho, J. E, Ruiz, A. L. T. G., Souza, K., Formagio, A. S. N. 2017. *Schinus terebinthifolius*: phenolic constituents and *in-vitro* antioxidant, antiproliferative and *in vivo* anti-inflammatory activities. *Brazilian Journal of Pharmacognosy*. 27: 445-452.
- Devasagayam, T. P. A, Tilak, J. C., Bloor, K. K., Sane, K. S., Ghaskadbi, S. S., and Lele, R. D. (2004). Free radicals and antioxidants in Human health: Current status and Future prospects. *Journal of the Association of Physician of India*. 52: 794-804
- Dhillon, H., Chikara, S., Reindl, K. M. (2014). Piperlongumine induces pancreatic cancer cell death by enhancing reactive oxygen species and DNA damage. *Toxicology Reports*, 1:309-318.
- Dixon, R. and Paiva, N. (1995). Stress-induced phenylpropanoid metabolism. *Plant Cell*, 17: 843-847
- Donepudi, M., Grutter, M. G. (2002). Structure and zymogen activation of caspases. *Biophysical Chemistry*. 101-102: 145-154
- Dragsted, L. O., Strube, M., & Larsen, J. C. (1993). Cancer-protective factors in fruits and vegetables: Biochemical and biological background. *Pharmacology and Toxicology*, 72, 116-135.
- Edwards, B. K., Ward, E., Kohler, B. A., Ehemann, C., Zaubers, A. G., Anderson, R. N. Annual report to the nation on the status of cancer, 1975-20006, featuring colorectal cancer trends and impact of interventions (risk factors, screening and treatment) to reduce future rates. Cancer.

- Ekaprasada, M. T. Nurdin, H, Ibrahim, S., and Dachriyanus. (2009). Antioxidant activity of methyl gallate isolated from leaves of *Toona sureni*, Indo. *J. Chem*, 9 (3): 457-460
- Elmore S. (2007). Apoptosis: A review of programmed cell death. *Toxicologic pathology*, 35(4), 495–516. doi:10.1080/01926230701320337
- Favaloro, B., Allocati, N., Graziano, V., Di Ilio, C., De Laurenzi, V. (2012). Role of Apoptosis in disease. *Aging*. 4(5):330-349
- Ferlay, J., Soerjomataram, I., Dikshit, R., Eser, S., Mathers, C., Rebelo, M., Bray, F., (2015). Cancer Incidence and mortality worldwide:sources, methods and major patterns in GLOBOCAN 2012. *Int J Cancer*, 136(5), E359-386. Doi:10.1002/ijc.29210
- Ferlay, J., Soerjomataram, I., Ervik, M., Dikshit, R., Eser, S., Mathers, C., Robelo, M., Parkin, D. M., Forman, D., Bray, F., GLOBOCAN 2012 v10, Cancer Incidence and Mortality Worldwide: IARC Cancer Base No. 11. Lyon, France: International Agency for Research on Cancer.
- Ferreira, M. L. F., Rius, S. P. and Casati, P. (2012). Flavonoids; biosynthesis, biological functions, and biotechnological applications. *Frontiers in Plant Science*, 3:1-15
- Fleming, M., Ravula, S., Tatishchev, S. F., and Wang, H. L. (2012). Colorectal carcinoma: Pathologic aspects. *Journal of Gastrointestinal Oncology*, 3(3); 153-173.
- Fontes, F., Severo, M. Castro, C., Lourenco, S., Gomes, S., Botelho, F., La Vecchia, C., Lunet, N., (2013). Model-based patterns in prostate cancer mortality worldwide. *British Journal of Cancer*, 108:2354-2366.
- Gilchrist, D. and Kosuge, T. (1980). Aromatic amino acid biosynthesis and its regulation. In BN Mifflin, ed, the Biochemistry of Plants, Academic Press, New York, 5: 507-531.
- Gobe, G., Rubin, M., Williams, G., Sawczuk, I., Buttyan, R. (2002). Apoptosis and expression of Bcl-2, Bcl-XL and Bax in renal cell carcinomas. *Cancer Investigation*. 20:324-332.
- Goldberg, A. A., Beach, A., Davies, G. F., Harkness, T. A., Leblanc, A., Titorenko, V. I. (2011). Lithocholic bile acid selectively kills neuroblastoma cells, while sparing normal neuronal cells. *Oncotarget*. 2:761-782.

- Gutzeit, H. O. and Ludwig-Muller, J. Biosynthesis and Chemical Properties of Natural Substances in Plants. In: *Plant Natural Products synthesis, Biological Functions and Practical Applications* (1st ed) .German: Wiley-VCH Verlag GmbH.
- Hail, N. (2005). Mitochondria: a novel target for the chemo prevention of cancer. *Journal of Apoptosis*. 10: 687-705
- Halliwell, B. (1997). Antioxidants and human disease: A general introduction. *Nutrition review*, 55; 44-52
- Hamilton, S., Vogelstein, B., Kudo, S., Riboli, E., Nakamura, S., Hainaut, P., Winawer, S. (2000). Tumors of the colon and rectum: carcinoma of the colon and rectum.
- Han, H., Landreneau, R. J, Santucci, T. S., Tung, M. Y., Macherey, R. S., Shackney, S. E., Sturgis, C. D., Raab, S. S., Silverman, J. F. Prognostic value of immunohistochemical expressions of p53, HER-2/neu, and bcl-2 in stage 1 non-small-cell lung cancer. *Human Pathology*. 33:105-110.
- Hassan, F. A., Ismail, A., Abdul, H. A., Azlan, A. (2011). Identification and quantification of phenolic compounds in Bambangan (*Mangifera pajang* Kost) peels and their free radical scavenging activity. *Journal of Agricultural and Food Chemistry*, 59(17):9102-11
- Hata, K., Hori, K., Takahashi, S. (2002). Differentiation- and apoptosis-inducing activities by pentacyclic triterpenes on a mouse melanoma cell line. *Journal of Natural Product*. 65(5): 645-64
- Hejmadi, M. (2009). *Introduction to cancer biology*: Bookboon.
- Hermann, K. M. (1995). The Shikimate pathway: Early steps in biosynthesis of aromatic compounds. *Plant Cell*, 7: 907-919.
- Hock, E. K., Prasad, K. N., Ismail, A., Mohd-Esa, N., (2010). Carotenoids from *Mangifera pajang*. And their antioxidant capacity. *Molecules*. 15: 6699-6712
- Hock, E. K., Azlan, A., Kin, W. K., Ismail, A., (2016). Phytochemicals and medicinal properties of indigenous tropical fruits with potential for commercial development. *Evidence-Based Complementary and Alternative Medicine*. 1-20
- Hollier, M., Whistler, T., Dowson, C., Vernon, S. D. (2007). Two optimized combination assays to examine apoptosis pathways in clinical samples. *Cytometry*. 71A: 675-685

- Ho, C. T., Osawa, T., Huang, M. T., Rosen, R. T. (1994). Food phytochemicals for cancer prevention II: Tea, Speces & Herbs. American Chemical Society, Washington, D. C.
- Hou, A. J., Peng, L. Y., Liu, Y. Z., Lin, Z. W., Sun, H. D. (2000). Gallotannins and related polyphenols from *Pistacia weinmannifolia*. *Plant Medicine*, 66: 624-626.
- Howlader, N., Noone, A. M., Krapcho, M., Miller, D., Bishop, K., Altekruse, S. F., Kosary, C. L., Yu, M., Ruhl, J., Tatalovich, Z., Mariotto, A., Lewis, D. R., Chen, H. S., Feuer, E. J., Cronin, K. A. (2016). SEER Cancer Statistics Review, 1975-2013. National Cancer Institute. Bethesda, MD. Available from: http://seer.cancer.gov/csr/1975_2013/
- Hsieh, T. J., Liu, T. Z., Chiu, C. C. (2004). Protective effect of methyl gallate from *Toona sinensis* (Meliaceae) against hydrogen peroxide-induced oxidative stress and DNA damage in MDCK cells. *Food and Chemical Toxicology*. 42(5):843-50.
- Ibrahim, M., Prasad, K. N., Ismail, A., Azlan, A., Abd Hamid, A., (2010). Physiochemical composition and antioxidant activities of underutilized *Mangifera pajang* fruit. *African Journal of Biotechnology*. 9(28): 4392-4397
- Ilic, M., and Ilic, I. (2016). Epidemiology of pancreatic cancer. *World Journal of Gastroenterol*, 22(44):9694-9705.
- Imperiale, T. F., Ransohoff, D. F., Itzkowitz, S. H., Levin, T. R., Lavin, P., Lidgard, G. P. (2014). Multitarget stool DNA testing for colorectal-cancer screening. *New England Journal of Medicine*, 370: 1287-97
- Inoue, M., Suzuki, R., Sakaguchi, N. Li, Z., Takeda, T., Ogihara, Y., Jiang, B. Y., and Chen, Y., (1995). Selective inductive of cell death in cancer cells by gallic acid. *Biological & Pharmaceutical Bulletin*, 18(11):1526-1530.
- Isuzugawa, K., Ogihara, Y., and Inoue, M. (2001). Different generation of inhibitors against gallic-acid induced apoptosis produce different sensitivity to gallic acid. *Biological Pharmaceutical Bulletin*, 24(3):249-253.
- Jemal, A., Siegel R, Xu J, Ward. (2010). Cancer Statistics 2010. *CA Cancer J Clin* 60: 277–300.
- Jess, T., Rungoe, C., and Peyrin-Biroulet, L. (2012). Risk of colorectal cancer in patients with Ulcerative Colitis: A Meta-analysis of Population-Based Cohort

- Studies. *Clinical Gastroenterology and Hepatology*, 10(6), 639-645. doi: <http://dx.doi.org/10.1016/j.cgh.2012.01.010>
- Kalra, N., Arora, A., Shukla, Y. (2006). Involvement of multiple signaling pathways in diallyl sulfide mediated apoptosis in mouse skin tumors. *Asian Pacific Journal of Cancer Prevention*. 7: 556.
- Kamatham, S., Kumar, N., and Gudipalli, P., (2015). Isolation and Characterization of gallic acid and methyl gallate from the seed coats of *Givotia rottleriformis* Griff. and their antiproliferative effect on human epidermoid carcinoma A431 cells. *Toxicology Reports* 2: 520-529
- Karim-Kos, H. E., de Vries, E., Soerjomataram, I., Lemmens, V., Siesling, S., Coebergh, J. W. (2008). Recent trends of cancer in Europe: A combined approach of incidence, survival and mortality for 17 cancer sites since the 1990s. *European Journal of Cancer*, 44:1345-1389
- Kasri, A. R. (1993). *Bul Epidemiol Kementerian Kesihatan Malaysia*. 3:3-24.
- Kaur, H, Inderjit & Kaushik, S. (2005). Cellular evidence of alleopathic interference of benzoic acid to mustard (*Brassica juncea* L.) seedling growth. *Plant Physiology and Biochemistry* 43: 77-81.
- Kaur, S., Michael, H., Arora, S., Harkonen, P. L., Kumar, S. (2005). The in-vitro cytotoxic and apoptotic activity of Triphala-an Indian Herbal drug. *Journal of Ethnopharmacology*, 97:15-20
- Kerr, J. F. R., Wyllie, A. H., Currie, A. R. (1972) Apoptosis: A basic biological phenomenon with wide-ranging implications in tissue kinetics. *Br. Journal of Cancer*. 26:239-257.
- Kerr, J. F. R., Winterford, C. M. Harmon, B. V. (1994). Apoptosis: Its significance in cancer and cancer therapy. *Cancer*. 73: 2013-2026
- Khoo, H. E., Prasad, K. N., Ismail, A., Mohd Esa, N. (2010). Carotenoids from *Mangifera pajang* and their antioxidant capacity. *Molecules*, 15(10):6699-712
- Knight, R. A., and Melino, G. (2011). Cell death in disease: from 2010 onwards. *Cell Death Dis*. 2:e202
- Kumar, P., Kumar, D. & Rai, K. N. (2016). Non-linear dual-phase-lag model for analyzing heat transfer phenomena in living tissues during thermal ablation. *Journal of Thermal Biology*, 60: 204-212.

- Ladas, E. J., Jacobson, J. S., Kennedy, D. D., Teel, K., Fleischauer, A., Kelly, K. M. (2004). Antioxidant and Cancer Therapy: A Systematic Review. *Journal of Clinical Oncology*. 22(3): 517-528.
- Lekha, P. K., Lonsane, B. K., (1997). Production and Application of Tannin Acyl Hydrolase: state of the art, *Advances in Applied Microbiology*, 44:215-260.
- Lakhani, S. R., Ellis, I. O., Schnitt, S. J. Tan, P. H. & Van de Vijver, M. J. (2012). *WHO Classification of Tumors of the Breast* (4th Revised edition ed). Lyon, France: International Agency for Research on Cancer.
- Lee, K. H., Lin, Y. M., Wu, T. S., Zhang, D. C. Yamagishi, T. Hayashi, T. Hall, I. H., Chang, J. J., Wu, R. Y and Yang, T. H. (1988). The cytotoxic principles of *Prunella vulgaris*, *Psychotria serpens*, and *Hyptis capotata*: Ursolic acid and related derivatives, *Planta Med*, 54(4): 308-311.
- Li, L., Dai, H. J., Ye, M., Wang, S. L., Xiao, X. J., Zheng, J., Chen, H. Y., Luo, Y., Liu, J. (2012). Lycorine induces cell-cycle arrest in the G0/G1 phase in K562 cells via HDAC inhibition. *Cancer Cell International*. 12;49.
- Lim, T. K. (2012). *Mangifera pajang* In: Edible Medicinal and Non-Medicinal Plants. Springer Netherlands: Springer Science and Bussiness Media BV: 131-134.
- Lodish, Berk & Zipursky. (2000). Section 24.1, Tumor Cells and the Onset of Cancer. *Molecular Cell Biology* (4th Edition. Ed). New York: W. H. Freeman.
- Liu, Q., Cao, Y., Zhou, P., Gui, S., Wu, X., Xia, Y., Tu, J. (2018). Pandurin inhibits cell proliferation by inducing G0/G1 phase cell cycle arrest and induces apoptosis in breast cancer cells. *Biomolecules and Therapeutics*. 26(3):328-334.
- Liu, F. S. (2009). Mechanism of Chemotherapeutic drug resistance in Cancer therapy- A Quick Review. *Taiwanese Journal of Obstetrics and Gynecology*. 48(3):239-244.
- Liu, R. H., (2003). Health benefits of fruits and vegetables are from additive and synergistic combination of phytochemicals. *The American Journal of Clinical Nutrition*. 78(suppl):517-520
- Mane, P. C., Kadam, D. D., Chaudari, R. D., Varpe, K. A., Shinde, R. S., Abhang, K. D., Sayyed, S. A. R. (2015). Phytochemical investigations of some green leafy vegetables for pharmacological importance. *Asian Journal of Pharmacology and Toxicology*. 3(8): 12-15.
- Mandal, S. M. Chakraborty, D., Dey, S. (2010). Phenolic acids act as signaling molecules in plant-microbe symbioses. *Plant Signal Behavior*, 5:359-368.

- Martin, S. J., Green, D. R. (1995). Protease activation during apoptosis: Death by thousand cuts? *Cell*. 82:349-352.
- Mayne, S. T., (1996). Beta-Carotene, carotenoids and disease prevention in human. *FASEB Journal*. 10:690-701.
- Mirfat, A. H., S., Salma, I., Razali, M., (2016). Natural antioxidant properties of selected wild *Mangifera* species in Malaysia. *Journal Tropical Agricultural and Foundation Science*. 44(1): 63-72
- Mohan, S., Abdul, A. B., Abdelwahab, S. I., Al-Zubairi, A. S., Sukari, M., A., Abdullah, R., Taha, M. M. E., Ibrahim, M. Y. & Syam, S. (2010). *Typhonium Flagelliforme* induces apoptosis in CEMss cells via activation of caspase-9, PARP cleavage and cytochrome c release: Its activation coupled with G₀ / G₁ phase cell cycle arrest. *Journal of Ethnopharmacology*, 131(3): 592-600.
- Mol, J., Grotewold, E., Koes, R. (1998). How genes paint flowers and seeds. *Trends in Plant Scice*, 3: 212-217
- Muhamad, M., Merriam, S., Suhami, N., (2012). Why Breast cancer patients seek traditional healers. *International Journal of Breast Cancer*. 2012, 1-9
- Mutalib, M. A., Ali, F. Othman, F., Ramasamy, R., & Rahmat, A. (2016). Phenolics profile and anti-proliferative activity of Cyphomandra Betacea fruit in breast and liver cancer cells. *SpringerPlus*, 5(1). Doi: 10.1186/s40064-016-3777-x
- Naczka, M. & Shahidi, F. (2006). Phenolics in cereals, fruits and vegetables: Occurrence, extraction and analysis. *Journal of Pharmaceutical and Biomedical Analysis*, 41 (5): 1523-1542
- National Academy of Sciences, National Research Council. (1982). Diet, Nutrition, and Cancer. National Academy Press, Washington, DC.
- Niemetz, R., Gross, G. G. (2005). Enzymology of gallotannin and ellagitannin biosynthesis. *Phytochemistry*, 66(17), 2001-2011.
- Omar Z. A., Ibrahim Tamin, N. S., (2011). National Cancer Registry Report; Malaysia Cancer Statistics- Data and Figure 2007, Ministry of Health Malaysia.
- Omar, Z., Ali, Z., Tamin, N. I. (2006). "Malaysian cancer and statistic- data and figure Peninsular Malaysia", in *National Cancer Registry 2006 Malaysia*. Pp. 1-112
- Orangi, M., Pasdaran, A., Shanehbandi, D., Kazemi, T., Yousefi, B., Hosseini, B. A., Baradaran, B. (2016). Cytotoxic and Apoptotic activities of Methanolic sub-fractions of *Scrophularia oxysepala* against Human Breast Cancer Cell line. *Evidence-based Complementary and Alternative Medicine*. 1-10

- Ouyang, D. I, Chen, J. J., Getzenberg, R. H., Schoen, R. E., (2005). Noninvasive testing for colorectal cancer; A review. *The American Journal of Gastroenterology*, 100; 1393-403
- Ozben, T. (2007). Oxidative stress and apoptosis: impact on cancer therapy. *Journal of Pharmaceutical Sciences*, 96:2181-2196.
- Pace, L. E., Keating, N. L., (2014). A systematic assessment of benefits and risks to guide breast cancer screening decisions. *Journal of American Medical Association*, 311: 1327-35.
- Paiva, P. M. G., Gomes, F. S., Napoleao, T. H., Sa, R. A., Correia, M. T. S., Coelho, L. C. B. B. (2010). Antimicrobial activity of secondary metabolites and lectins from plants. *Current research technology and Education Topics in Applied Microbiology and Microbial Biotechnology*, pp. 396-406.
- Park, C., Moon, D. O., Rhu, C. H., Choi, B. T., Lee, W. H., Kim, G. Y., Choi, Y. H. (2007). B-Sitosterol induces anti-proliferation and apoptosis in human leukemic U937 Cells through activation of caspase-3 and induction of Bax/Bcl ratio. *Biological and Pharmaceutical Bulletin*, 30:1317-1323
- Parkin, D. M., Bray, F., Ferlay, J., Jemal, A. (2014). Cancer in Africa. *Cancer Epidemiology, Biomarkers and Prevention*, 23:953-966.
- Parkin, D. M., Boyd, L., Walker, L. C. (2011). The fraction of cancer attributable to lifestyle and environmental factors in the UK in 2010. *British Journal of Cancer*. 105: S77-S81.
- Parsaeimehr, A., Sargsyan, E., Vardanyan, A. (2011). Expression of secondary metabolites in plants and their useful perspective in animal health. *International Journal of the Bioflux Society*, 3(2): 115-124
- Partridge, A. H., Burstein, H. J., Winer, E. P., (2001). Side effects of chemotherapy and combined chemohormonal therapy in women with early stage of breast cancer. *Journal of National Cancer Institute Monographs*. 30:135-142.
- Patwardhan, B. *Drug discovery and Development. Traditional medicine and Ethnopharmacology*. Page 32. New India Publishing Agency. pp 32. 2007
- Placzek, W. J., Wei, J. Kitada, S., Zhai, D., Reed, J. C., Pellechia, M. A survey of the anti-apoptotic Bcl-2 subfamily expression in cancer types provides a platform to predict the efficacy of Bcl-2 antagonists in cancer therapy. *Cell Death Dis*.1:e40

- Prasad, K. N., Hassan, F. A., Yang, B., Kong, K. W., Ramanan, R. N., Azlan, N. (2011). Response surface optimization for the extraction of phenolic compounds and antioxidant capacities of underutilized *Mangifera pajang* Kosterm. Peels. *Food Chemistry*, 128(4):1121-7
- Ramulu, P., Rao, P. U., (2003). Total, insoluble and soluble dietary fiber contents of Indian fruits. *Journal of Food Composition. Anal.* 16:677-685.
- Ravishankar, D., Rajora, A. K. Greco, F., Osborn, H. M. I. (2013). Flavonoids as prospective compounds for anti-cancer therapy. *The International Journal of Biochemistry & Cell Biology* 45, 2821-2931.
- Redwood, D., Provost, E., Asay, E., Roberts, D., Haverkamp, D., Perdue, D. (2014). Comparison of fecal occult blood tests for colorectal cancer screening in an Alaska Native population with high prevalence of *Helicobacter pylori* infection, 2008-2012. *Preventing Chronic Disease*, 11; E56.
- Reece, J. B., Urry, L. A., Cain, M. L., Wasserman, S. A., Minorsky, P. V., Jackson, R. B., (2011). *The Cell Cycle*. Campbell Biology: Pearson Inc.
- Rohmer, M. (1999). The discovery of mevalonate-independent pathway for isoprenoid biosynthesis in bacteria, algae and higher plants. *National Products Reports*. 16: 565-74
- Rukayah, A. (2002). *Buah-buahan Malaysia*. Kuala Lumpur: Dewan Bahasa dan Pustaka
- The Plant List. 2010. Version 1. Published on the Internet. <http://www.theplantlist.org/> (accessed 28 May 2016)
- Ryan, K. G. Swinny, E. E., Markham, K. R., Winefield, C. (2002). Flavonoid gene expression and UV photoprotection in transgenic and mutant *Petunia* leaves. *Phytochemistry*, 59:23-32
- Saleem, A, Husheem, M., Harkonen, P., and Pihlaja, K. (2002). Inhibition of Cancer growth by crude extract and the phenolics of *Terminalia chebula* retz.fruit. *Journal of Ethnopharmacology*. 81:327-336.
- Saraste, A., Pulkki, K. (2000). Morphologic and biochemical hallmarks of apoptosis. *Cardiovascular Research*. 45(3):528-537.
- Schuurink, R. C., Haring, M. A. and Clark, D. G. (2006). Regulation of volatile benzenoid biosynthesis in *petunia* flowers. *Trends in Plant Science*, 11:20-25.
- Sewda, K., Coppola, D., Enkemann, S., Yue, B., Kim, J., Lopez, A. S., Morse, D. L. (2016). Cell surface markers for colon adenoma and adenocarcinoma. *Oncotarget*, 7(14), 17773-17789. doi:10.18632/oncotarget.7402

- Song, X., Zhang, Y., Wang, X., Zhang, W., Wang, Z., Zhang, F., Gu, J. (2017). Casticin induces apoptosis and G0/G1 cell cycle arrest in gallbladder cancer cells. *Cancer Cell International*, 17, 9.
- Stewart, B. W., & Wild, C., (2014). World Cancer Report 2014. International Agency for Research on Cancer. *World Health Organization*, 505.
- Subramaniam, A. P., John, A. A., Vellayappan, M., V., Balaji, A., Jaganathan, S. K., Supriyanto, E., and Yusof, M. (2015). Gallic acid: prospects and molecular mechanisms of its anticancer activity., *Royal Society of Chemistry Advance*, 5, 35608-35621.
- Sun, S. Y., Hail Jr, N., Lotan, R. (2004). Apoptosis as a novel target for cancer chemo prevention. *Journal of the National Cancer Institute*. 96: 662-672.
- Tangah, J., Bajau, F. E., Jilimin, W., Chan, H. T., Wong, S. K., Chan, E. W. C. (2017). Phytochemistry and Pharmacology of *Mangifera pajang*: An Iconic fruit of Sabah, Malaysia. *Systemic Reviews in Pharmacy*. 8(1) :86-91.
- Tamimi, R. M., Lagiou, P., Adami, H. O and trichopoulos, D. (2002). Prospects for chemiprevention of cancer. *Journal of Internal Medicine*. 251, 286-300.
- Tawfik, K., Kimler, B. F., Davis, M. K., Fan, F, Tawfik, O. Prognostic significance of Bcl-2 in invasive mammary carcinomas; a comparative clinicopathologic study between 'triple-negative' and 'non-triple negative' tumors. *Human Pathology*.
- Tolonen, A., (2003). Analysis of secondary metabolites in plant and cell culture tissue of *Hypericum perforatum* L. and *Rhodiola rosea* L. An Academic Disertation from Acta Universitatis Oulensis Scientiae Rerum Naturalium (<http://herkules oulu.fi/issn 03553191/>)
- Torres-Leon, C., Ventura-Sobrevilla, J., Serna-Cock, L., Ascacio-Valdes, J. A., Contreras-Esquivel, J., Aguilar, C. N. (2017). Pentagalloylglucose (PGG): A valuable phenolic compound with functional properties. *Journal of Functional Food*. 37: 176-189.
- Taylor, D. P., Burt, R. W., Williams, M. S., Haug, P. J. and Cannon-Albright, L. A. (2010). Population-Based Family History-Specific risks for colorectal cancer: A constellation Approach. *Gastroenterology*, 138(3), 877-885.
- Thornberry, N. A., Lazebnik, Y. (1998). Caspases: enemies within *Science*. 281: 1312-1316.
- Thompson, R. J., Jacques, D., Haslam, E., and Tanner, R. J. N. (1972). Plant proanthocyanidins. Introduction the isolation, structure and distribution in

- nature of plant procyanidins. *Journal of Chemical Society Perkins Transaction*, 1:1387
- Torre, L. A., Siegel, R. L., Ward, E. M., and Jemal, A. (2015). Global cancer Incidence and Mortality Rates and Trends- An update. *Cancer Epidemiol Biomarkers Prev*, 25(1), 16-27. Doi: 10.1158/1055-9965.epi-15-0578
- Tzin, V. and Galili, G. (2010). The biosynthesis pathways for Shikimate and Aromatic amino acids in *Arabidopsis thaliana*. *American Society of Plant Biologists*: 1-18.
- Vainio, H. Bianchini, F., (2002). Editors, Breast cancer screening (volume 7). Lyon: International Agency for Research on Cancer Press.
- Van De Staaiji, J., De Bakker, N. V., Oosthoek, A., Broekman, R., Van Beem, A., Stroetenga, M., Aerts, R. Rozema, J. (2002). Flavonoids concentrations in three grass species and a sedge grown in the field and under controlled environment conditions in response to enhanced UV-B radiation. *Journal of Phytochemistry and Photobiology*, 66:21-29.
- Vita, J. A. (2005). Polyphenol and cardiovascular disease: effect on endothelial and platelet function. *American Journal of Clinical Nutrition*, 81; 292-297.
- Verdonk, J. C., Ric de Vos, C. H., Verhoeven, H. A., Haring, M. A., van Tunen, A. J., and Schuurink, R. C. (2003). Regulation of floral scent production in petunia revealed by targeted metabolomics. *Phytochem*, 62:997-1008.
- Vermes, I., Haanen, C., Steffens-Nakken, H., Reutellingsperger, C. (1995). A novel assay for apoptosis Flow cytometric detection of phosphatidylserine expression on early apoptotic cells fluorescein labelled Annexin V. *Journal of Immunological Methods*. 184(1): 39-51.
- Viard-Leveugle, I, Veyrenc, S., French, L. E., Brambilla, C., Brambilla, E. Frequent loss of Fas expression and function in human lung tumors with overexpression of FasL in small cell lung carcinoma. *Journal of Pathology*. 201: 268-277.
- Vogt, T. (2010). Phenylpropanoid biosynthesis. *Mol Plant*, 3:2-20.
- Wang, H., Provan, G. J., Helliwell, K., (2003). Determination of hamamelitannin, catechins and gallic acid in which hazel bark, twig and leaf by HPLC. *Journal of Pharmaceutical Biomed. Anal*, 33(4), 539-544
- Weiss, W., Benarde, M. A. (1983). The temporal relation between cigarette smoking and pancreatic cancer. *American Journal Public Health*, 73:1403-1404.

- Wildermuth, M., Dewdney, J., Wu, G. and Ausubel, F. (2001). Isochorismate synthase is required to synthesize salicylic acid for plant defence. *Nature*, 414: 562-565.
- Williams, R., Spencer, J., Rice-Evans, C. (2004). Flavonoids: Antioxidants or signaling molecules. *Free Radical Biology Medicine*, 36:838-849.
- Williamson, P., Eijnde, S., Schlegel, R. A. (2001). Chapter 15 Phosphatidylserine exposure and phagocytosis of apoptotic cells. *Method in Cell Biology*. 66: 339-364
- Willet, W. C. (2000). Diet and Cancer. *Oncologist*, 5:393-404.
- Willet, W. C. (1995). Diet, nutrition, and avoidable cancer. *Environmental Health Perspectives*, 103 (8):165-170.
- World Conservation Monitoring Centre. (1998). *Mangifera pajang*. IUCN Red List of Threatened Species 1998:e T31394A9625586. <http://dx.doi.org/10.2305/IUCN.UK.1998.RLTS.T31394A9625586.en>. Downloaded on 04 October 2018.
- Wong, K. C., Siew, S. S. (1994). Volatile components of the fruits of Bambangan (*Mangifera pajang* Kostermans) and Binjai (*Mangifera caesia* Jack). *Flavor and Fragrance Journal*. 9:173-178.
- Wong, C. C., Li, H. B., Cheng, K.W. and Chen, F. (2006). A systematic survey of antioxidant activity of 30 Chinese medicinal plants using ferric reducing antioxidant power assay. *Food Chemistry*, 97(4): 705-711
- Wong, A. S., Che, C. M. & Leung, K. W. (2015). Recent advances in ginseng as cancer therapeutics: A functional and mechanistic overview. *The Royal Society of Chemistry*.
- Wong, M. C. S., Goggins, W. B., Wang, H. H. X., Fung, F. D. H., Leung, C., Wong, S. Y. S., Ng, C. F., Sung, J. J. Y. (2016). Global Incidence and mortality for prostate cancer: Analysis of temporal patterns and trends in 36 countries. *European Urology*, 70(5): 862-874.
- Wong, M. C. S., Jiang, J. Y., Liang, M., Fang, Y., Yeung, M. S. and Sung, J. J. Y. (2017). Global temporal patterns of pancreatic cancer and association with socioeconomic development. *Scientific Reports*, 7:1-9. doi: 10.1038/s41598-017-02997-2
- Wu, Q. J., Wu, L., Zheng, L. Q., Xu, X., Ji, C., Gong, T. T. (2016). Consumption of fruits and vegetables reduces risk of pancreatic cancer: Evidence from epidemiological studies. *European Journal of Cancer Prevention*, 25: 196-205.

- Yip, C. H., Pathy, N. B., Teo, S. H., (2014). A review of Breast Cancer Research in Malaysia. *Medical Journal Malaysia*. 69: 1-15.
- You, B. R, and Park, W. H., (2010). Gallic-acid induced lung cancer is related to glutathione depletion as well as reactive oxygen species increase. *Toxicol In Vitro*, 24: 1356-1362.
- Yuan, G. Q., Li, Q. Q., Qin, J. (2012). Isolation of methyl gallate from *Toxicodendron sylvestre* and its effect on tomato bacterial wilt. *Plant disease*. 96(8):1143-1147.
- Zabidah, A. A., King, K. W. & Amin, I., (2011). Antioxidant properties of tropical juices and their effects on in-vitro haemoglobin and low-density lipoprotein (LDL) oxidation. *International Food Research Journal*, 18:549-556.
- Zelen, M. (1988). Are primary cancer prevention trials feasible? *Journal of the National Cancer Institute*. 80(18): 1442-1444.
- Zhang, J., Dhakal, I. B., Zhao, Z., Li, L. (2012) Trends in mortality from cancers of the breast, colon, prostate, esophagus and stomach in East Asia: role of nutrition transition. *European Journal of Cancer Prevention*, 21:480-9
- Zhang, J., Zhao, Z., Berkel, H. J. (2005). Animal fat consumption and pancreatic cancer incidence: Evidence of interaction with cigarette smoking. *Annals of Epidemiology*, 15:500-508.
- Zhou, Y., Li, Y., Zhou, T., Zheng, J. Li, S. and Li, H. B. (2016). Dietary Natural Products for prevention and treatment of liver cancer. *Nutrients*, 8(3), 156. doi : 10.3390/nu8030156
- Zima, T., Fialova, L., Mastek, O. Janebova, M., Crkovka, J., Malbohan, I., Popov, P. (2001). Oxidative stress, metabolism of ethanol and alcohol-related diseases. *Journal of Biomedical Science*, 8, 59-70.
- Zouhiri, F., Mouscadet, J. F., Mekouar, K., Desmaele, K., Savoure, D., Leh, D., Subra, H., Bret, F., Auclair, C and d'Angelo, J. (2000). Structure-based activity relationships and binding mode of styrylquinolines as potent inhibitors of HIV-1 integrase and replication of HIV in cell culture. *Journal of Medical Chemistry*, 43(8):1533-1540.